

# Wash Buffer I and II

SKU: WASH1-004L and WASH2-004L

Size: 4L per Cubitainer

## Intended Use:

For Research Use Only

## Application:

Post-hybridization washing of fluorescence in situ hybridization (FISH) and chromogenic in situ hybridization (CISH) slides.

## Composition:

Component	Concentration	Volume
FISH Wash Buffer I	0.4x SSC, 0.3% IGEPAL	4L
FISH Wash Buffer II	2x SSC, 0.1% IGEPAL	4L

## Storage, Handling, Shelf Life, and Disposal:

Store product at 15-30°C; avoid frequent fluctuations in temperature. Expiration date noted on product. Gloves and other protective equipment should be used when handling the Wash Buffers. Dispose as per local regulations.

## Warning and Precaution:

Product does not contain any human or animal components. See Safety Data Sheet (SDS) for more detailed safety and handling information. Do not use expired buffers. Avoid cross-contamination. Read instructions for use in full before use.

## Limitations:

This product is for research use only. Performance is dependent on sample preparation, sample quality, and proper storage. This product is only for use by trained laboratory professionals.

## Contact and Support:

3370 Walden Avenue, Suite 114  
Depew, New York, 14043  
Phone: (716) 856-3873 or 1-800-715-5880  
Fax: (716) 431-3440  
info@empiregenomics.com  
www.empiregenomics.com

## Reagent(s) Not Provided:

- DAPI with Antifade

## Equipment Not Provided:

- Coplin Jars
- Water Bath
- 22x22mm Glass Coverslip
- Lint-Free Wipe
- Timer
- Pipettes (P10)
- Minifuge
- Vortex

## FISH Post-Hybridization Wash Protocol:

1. Place a coplin jar filled with Wash Buffer I in a 72°C water bath. Allow the buffer to come to temperature.
2. Place a coplin jar filled with Wash Buffer II at room temperature.
3. Remove the coverslip from the slide.
4. Aggressively agitate the slide in the coplin jar containing Wash Buffer I for ~10 seconds. Allow the slide to sit in the coplin jar for two minutes. When removing the slide, aggressively agitate the slide again for ~10 seconds.
5. Aggressively agitate the slide in the coplin jar containing Wash Buffer II for ~10 seconds. Allow the slide to sit in the coplin jar for two minutes. When removing the slide, aggressively agitate the slide again for ~10 seconds.
6. Wipe the back of the slide with a lint-free wipe.
7. Allow the slide to air-dry in the dark for 15 minutes.
8. Once dry, apply 10uL of DAPI with Antifade per cellular area and cover with a 22x22mm glass coverslip.
9. Place the slides in the dark at -20°C for 10-20 minutes before viewing under the microscope.
10. Store washed slides at -20°C in the dark.